NO: gas; BP -151.7 °C; vapor density

VISIBLE ABSORPTION

**SPECTROPHOTOMETRY** 

standard solutions of NO 2

3 to 18 µg NO<sub>2</sub> per sample [1]

NO: 0.061 [1]; NO<sub>2</sub>: 0.026 [2]

nitrite ion, NO2

540 nm

EXTRACTION SOLUTION: absorbing solution, 50 mL

NO<sub>2</sub>: gas; MP -11.2 °C; BP 21 °C;

vapor density (air=1) 2.83

(air=1) 1.0

# NITRIC OXIDE and NITROGEN DIOXIDE

NO MW: 30.01 CAS: 10102-43-9 RTECS: QX0525000 46.01 10102-44-0 QW9800000 NO<sub>2</sub>

METHOD: 6014, Issue 1 **EVALUATION: FULL** Issue 1: 15 August 1994

**PROPERTIES:** 

TECHNIQUE:

ANALYTE:

**WAVELENGTH:** 

**CALIBRATION:** 

PRECISION (S.):

OSHA: 25 ppm NO; C 1 ppm NO,

NIOSH: 25 ppm NO; 1 ppm STEL NO,

ACGIH: 25 ppm NO; 3 ppm TWA, 5 ppm STEL NO,

 $(1 \text{ ppm NO} = 1.227 \text{ mg/m}^3 @ \text{NTP})$  $(1 \text{ ppm NO}_2 = 1.882 \text{ mg/m}^3 @ \text{NTP})$ 

SYNONYMS: NO: nitrogen monoxide

NO2: nitrogen peroxide; nitrogen tetroxide

**SAMPLING MEASUREMENT** 

SAMPLER: SORBENT TUBES

(oxidizer + 2 triethanolamine-treated

molecular sieve)

FLOW RATE: NO: 0.025 L/min

NO<sub>2</sub>: 0.025 - 0.2 L/min

VOL-MIN: 1.5 L

-MAX: 6 L

SHIPMENT: routine

SAMPLE

STABILITY: stable at least 7 days @ 25 °C [1,2]

**BLANKS:** 3 to 6 field blanks and 10 media blanks pe

set

**ESTIMATED LOD:** 1 μg NO<sub>2</sub> per sample [3]

RANGE:

**ACCURACY** 

**RANGE STUDIED:** NO: 11-48 ppm [1]; NO  $_2$ : 2-12 ppm [2]

(1.5-L samples) (3-L samples)

NO: 4.1% [1]; NO<sub>2</sub>: -2% [2] **OVERALL PRECISION (Ŝ<sub>rT</sub>):** NO:0.083 [1]; NO<sub>2</sub>:0.063 [2]

**ACCURACY:** NO: ± 20.4%;

 $NO_2$ : ± 14.6%

APPLICABILITY: The working range for NO is 1 to 50 ppm (1.3 to 61 mg/m<sup>3</sup>) for a 1.5-L air sample. The working range for NO <sub>2</sub> is 0.5 to 25 ppm (1 to 47 mg/m<sup>3</sup>) for a 3-L air sample. The lower sampling rate for NO is to allow collection of oxidized NO on the back sorbent section. At the lower rate, both NO and NO 2 may be determined simultaneously.

INTERFERENCES: Any compound that reacts with the colorimetric reagents will interfere.

OTHER METHODS: This method is based on that of Willey, et al. [4] and combines S321, S320, and P&CAM 231 in a revised format [3,5]. OSHA methods ID-182 and ID-190 use the same sampler, with measurement by ion chomatography [6].

### **REAGENTS:**

- 1. Triethanolamine, TEA, reagent grade.
- 2. n-Butanol, reagent grade.
- 3. Phosphoric acid, H<sub>3</sub>PO<sub>4</sub>, conc., reagent grade.\*
- <u>N</u>-(1-napthyl)ethylenediaminedihydrochloride, NEDA.
- 5. Sodium nitrite, NaNO 2.
- Absorbing solution: Dissolve 15.0 g triethanolamine in ca. 500 mL deionized water, add 0.5 mL n-butanol, and dilute to 1 L.
- 7.  $H_2O_2$  solution, 0.02% (v/v): Dilute 0.2 mL of 30%  $H_2O_2$  to 250 mL with deionized water.
- 8. Sulfanilamide solution: Dissolve 10 g sulfanilamide in 400 mL deionized water, add 25 mL conc. H <sub>3</sub>PO<sub>4</sub>, and dilute to 500 mL.
- NEDA solution: Dissolve 0.5 g <u>N</u>-(1-napthyl) ethylenediamine dihydrochloride in 500 mL deionized water.
- Calibration stock solution, 100 NO <sup>2</sup> μg/mL: Dissolve 0.1500 g NaNO <sup>2</sup> in 1 L deionized water.
  - \* See SPECIAL PRECAUTIONS.

#### **EQUIPMENT:**

 Sampler: Three glass tubes, 7-mm OD, flamesealed ends with plastic caps, with glass wool retainers:

Tube A: 400 mg TEA-coated molecular sieve (type 13x, 30-40 mesh)

Tube B: 800 mg oxidizer (chromate) to convert NO to NO 2.

Tube C: Same as Tube A.

Connect the tubes in series with flexible tubing. Position Tube C closest to the inlet of the sampling pump. Tubes are commercially available (SKC-226-40, or equivalent).

- 2. Personal sampling pump, 0.025 to 0.2 L/min, with flexible connecting tubing.
- 3. Spectrophotometer, UV-visible (540 nm), with cuvettes, 1-cm silica cuvettes.
- 4. Beakers, borosilicate, 100-mL.
- 5. Volumetric flasks, 50-mL and other convenient sizes
- 6. Pipets, 1-, 5-, 10-mL and other convenient sizes.
- 7. Stopwatch.

**SPECIAL PRECAUTIONS:** Concentrated acid is corrosive to the skin and mucous membranes. Handle it only in a hood.

#### SAMPLING:

- 1. Calibrate the sampling pump with a representative sampler in line.
- 2. Immediately before sampling, break ends of sampler and attach to pump.

NOTE: Nitrogen dioxide collects on the first tube (Tube A), and is thereby separated from nitric oxide, which is oxidized by Tube B and then is collected on Tube C (adjacent to the sampling pump.)

- 3. Sample at an accurately known flow rate of 0.025 L/min  $\pm$  5%.
  - NOTE: If nitric oxide is not to be determined, a flow rate of up to 0.2 L/min may be used.
- 4. Cap the sampler and pack securely for shipment. Submit adequate numbers of field blanks and media blanks to the laboratory.

## **SAMPLE PREPARATION:**

- 5. Transfer the sorbent from Tube A and Tube C to separate 50-mL volumetric flasks. Discard glass wool plugs and oxidizer (Tube B).
- 6. Add absorbing solution to sample in 50-mL volumetric and bring to the mark.
- 7. Stopper flask and shake vigorously for 30 sec. Allow solids to settle.
- 8. Pipet 10 mL of extracted sample into a 50-mL volumetric flask. NOTE: Start reagent blanks at this step.
- 9. Add 1.0 mL hydrogen peroxide solution, 10.0 mL sulfanilamide solution, and 1.4 mL NEDA solution. Mix thoroughly after each addition.
- 10. Allow 10 min for complete color development.

#### **CALIBRATION AND QUALITY CONTROL:**

- 11. Calibrate daily with at least six working standards to cover the range of 1 to 18 μg nitrite ion per 10-mL sample.
  - a. Analyze the working standards together with blanks and samples (steps 8 through 10 and steps 12 through 14).
  - b. Prepare a calibration graph [absorbance vs. μg NO <sup>2</sup> per sample].

#### **MEASUREMENT:**

- 12. Set wavelength on the spectrophotometer to 540 nm.
- 13. Set to zero with reagent blank.
- 14. Transfer some of the sample solution from step 10 to a cuvette and record the absorbance.

## **CALCULATIONS:**

- 15. From the calibration graph, determine the mass of NO  $_2$  in each Tube A, W  $_A$  (µg), and in the corresponding average blank, B  $_A$  (µg). Similarly, determine the mass of NO  $_2$  in each Tube C, W  $_c$  (µg), and average blank, B  $_c$  (µg).
- 16. Calculate the concentration, C <sub>NO2</sub> (mg/m³) of NO<sub>2</sub> in the volume of air sampled, V (L), applying the conversion factor 0.63:

$$C_{NO_2} = \frac{(W_A - B_A)}{0.63 \text{ V}}, \text{ mg/m}^3.$$

NOTE: The conversion factor 0.63 represents the number of moles of nitrite ion produced by 1 mole of nitrogen dioxide gas. For NO or NO <sub>2</sub>, gas concentrations above 10 ppm, use 0.5 as the conversion factor [7].

17. Calculate the concentration, C <sub>NO</sub> (mg/m³), of NO in the air volume sampled, V (L), applying the factor 0.652 (MW NO/MW NO <sub>2</sub>) and the conversion factor 0.63:

$$C_{NO} = \frac{(W_c - B_c) \cdot 0.652}{0.63 \text{ V}}, \text{ mg/m}^3.$$

### **EVALUATION OF METHOD:**

Method S321, Nitric Oxide, was evaluated over the range of 11.1 to 47.7 ppm (13.8 to 58.5 mg/m <sup>3</sup>) for 1.5-L samples, collected from dynamically generated test atmospheres [1,8]. The test concentration was verified with a direct reading instrument, Energetic Sciences Enolyzer. The 1.2 g oxidizer section was found adequate for a 60-min sampling time. NO samples had a mean recovery of 99.5% after 7 days storage at ambient temperature.

Method S320, Nitrogen Dioxide, was evaluated over the range 3.0 - 11.6 ppm (5.8 to 21.6 mg/m  $^{3}$ ) using 3.9-L samples [2,7]. When an atmosphere at 84% RH containing 11.59 ppm NO  $_{2}$  was sampled at 0.064 L/min, 1.0% breakthrough occurred after 60 min and 2.4% breakthrough occurred after 180 min. Quantitative recovery was obtained for samples containing 47  $\mu$ g NO  $_{2}$  which were stored for 12 days at ambient conditions.

### **REFERENCES:**

- [1] Backup Data Report for Nitric Oxide, S321, prepared under NIOSH Contract No. 210-76-0123.
- [2] Backup Data Report for Nitrogen Dioxide, S320, prepared under NIOSH Contract No. 210-76-0123.
- [3] NIOSH Manual of Analytical Methods, 2nd ed., Vol. 4, Methods S320 and S321. U.S. Department of Health, Education, and Welfare. DHEW (NIOSH) Publication No. 78-175.
- [4] Willey, M.A., C.S. McCammon, and L.J. Doemeny, Am. Ind. Hyq. Assoc. J. 38, 358 (1977).
- [5] NIOSH Manual of Analytical Methods, 2nd ed., Vol. 1, P&CAM 231, U.S. Department of Health, Education, and Welfare (NIOSH) Publ. 77-157-A (1977).
- [6] OSHA Analytical Methods Manual, 2nd ed., Part 2, Vol. 2, ID-182 and ID-190, U.S. Department of Labor, Salt Lake City, UT (1991).
- [7] Gold, A., Anal. Chem. 49, 1443-1450 (1977).
- [8] NIOSH Research Report Development and Validation of Methods for Sampling and Analysis of Workplace Toxic Substances, U.S. Department of Health and Human Services, Publ. (NIOSH) 80-133 (1980).

## **METHOD REVISED BY:**

W. J. Woodfin and M. E. Cassinelli, NIOSH/DPSE; Method S321 validated under NIOSH Contract No. 210-76-0123.